

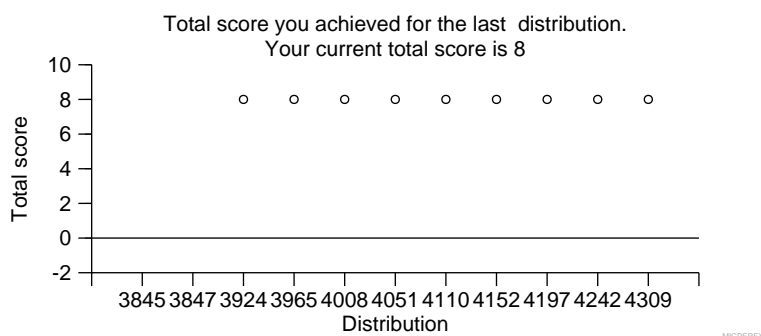
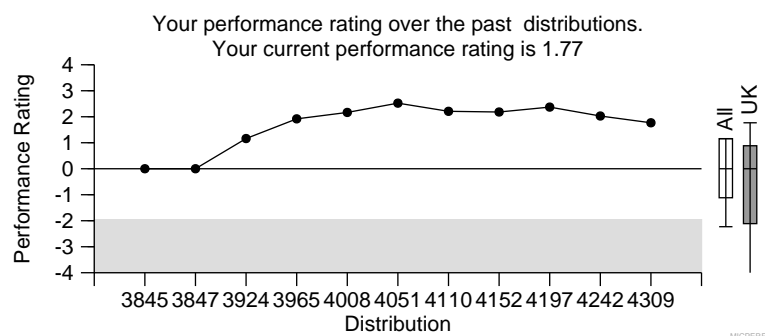
Intended Result	Your Report	Your Score
Specimen 4463 <i>Plasmodium falciparum</i> nucleic acid present	<i>Plasmodium falciparum</i> nucleic acid present	2
Specimen 4464 No <i>Plasmodium</i> nucleic acid detected	No <i>Plasmodium</i> nucleic acid detected	2
Specimen 4465 <i>Plasmodium vivax</i> nucleic acid present	<i>Plasmodium vivax</i> nucleic acid present	2
Specimen 4466 <i>Plasmodium ovale</i> nucleic acid present	<i>Plasmodium ovale</i> nucleic acid present	2

Cumulative score information

Total number of specimens sent to you for **UK NEQAS for Malaria (molecular)** over the last 2 distributions is 8.
For these distributions specimen numbers 4260 4261 4262 4263 4463 4464 4465 4466 have been analysed and scored.
Number of reports analysed 8
Number of specimens reported as not examined (not scored) 0
Number of specimens received too late for analysis (not scored) 0
Number of specimens for which no report was received (not scored) 0
Your cumulative score for these specimens was 16 out of a possible total of 16

Performance rating

Your performance rating for **UK NEQAS for Malaria (molecular)** (i.e. the number of standard errors by which your cumulative score lies above or below the mean) for **UK** laboratories is 1.77.
A performance rating of more than 1.96 standard errors below the mean indicates possible poor performance.
Please note your performance rating may alter if other participants' results are amended.
No score penalty is incurred for non return of reports. However non return of results may be used as a measure of poor performance.



Turn around time: The time taken to report your results was 0 day(s). This information is provided for your own use and does not form part of your performance assessment.

Comments

Results obtained by participants were in good agreement with the pre- and post distribution results obtained at the UK NEQAS Parasitology lab. Those participants who reported discrepant results are reminded to follow the manufacturers' instructions and to investigate the root cause of the discrepancy.

The figures in the histograms and those in the overall results tables may differ (1) due to exclusion of kits displayed in the histograms resulting in apparently lower numbers of data sets in the histograms or (2) due to participants using more than one kit resulting in higher numbers of data sets in the histograms.

Participants are reminded that the intended results are available on the UK NEQAS website usually on the day following the closing date.

Repeat specimens

Repeat specimens can be supplied on request from organiser@ukneqasmicro.org.uk and participants are asked to request these, quoting their laboratory number, as soon as the intended results are displayed on the web or on receipt of their report.

Enquiries

Telephone and written enquiries can be made to Dr. Jaya Shrivastava : +44(0)20 39081371

E-mail address: organiser@ukneqasmicro.org.uk

Participants who perform only genus identification and/or do not perform identification of *P. knowlesi* are requested to inform us so they can be scored accordingly.

This report was authorised by Professor P L Chiodini, Parasitology Scheme Organiser and Dr Jaya Shrivastava, Parasitology Scheme Manager.

Participants are reminded to quote their Laboratory ID number in all correspondences.

We thank colleagues in the PHE National Parasitology Reference Laboratory (Dr Spencer Polley) and the PHE Malaria Reference Laboratory (Professor Colin Sutherland and Dr Debbie Nolder) for undertaking polymerase chain reaction for confirmation of malaria species for this scheme.

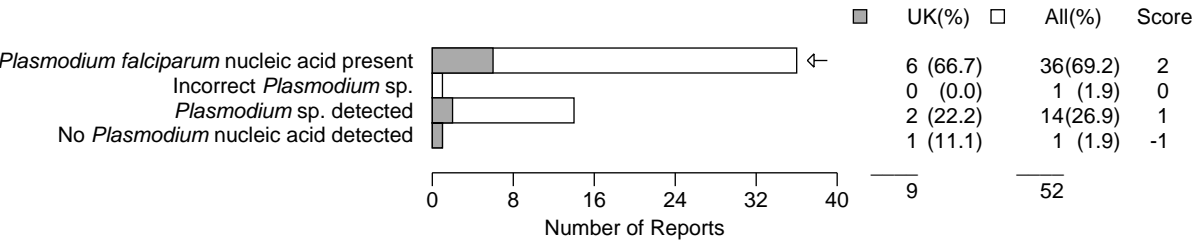


All specimens in this distribution were from a single patient sample and were diluted to the required parasite density using a donor blood which was negative for malaria DNA.

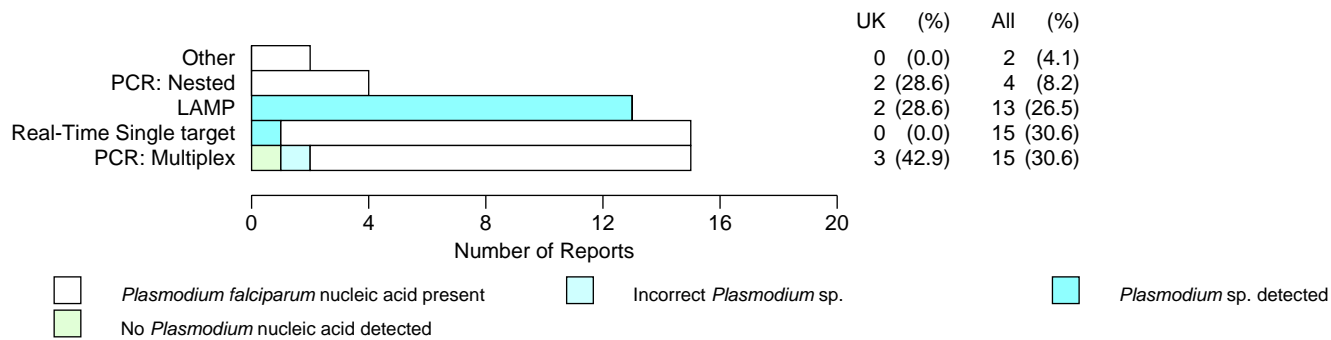
All samples were examined pre- and post- distribution by Nested PCR (modified Snounou protocol adapted from Snounou et al; 1993) and gave the intended results.

Please note that we report the parasitaemia in parasites/mL. Participants wanting to know the parasitaemia in parasites/uL are advised to divide the figure by 1000.

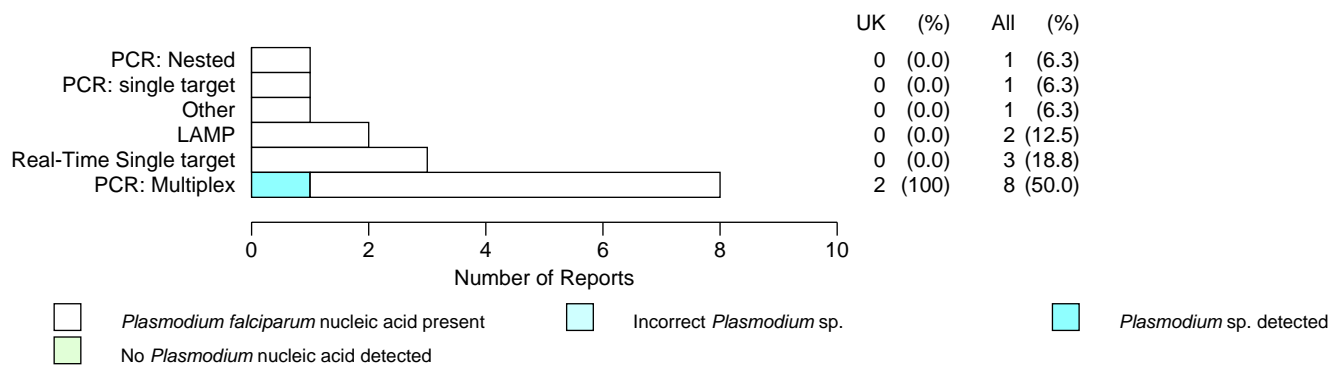
Specimen : 4463 *Plasmodium falciparum*: 200,000 parasites/mL **Overall Results**



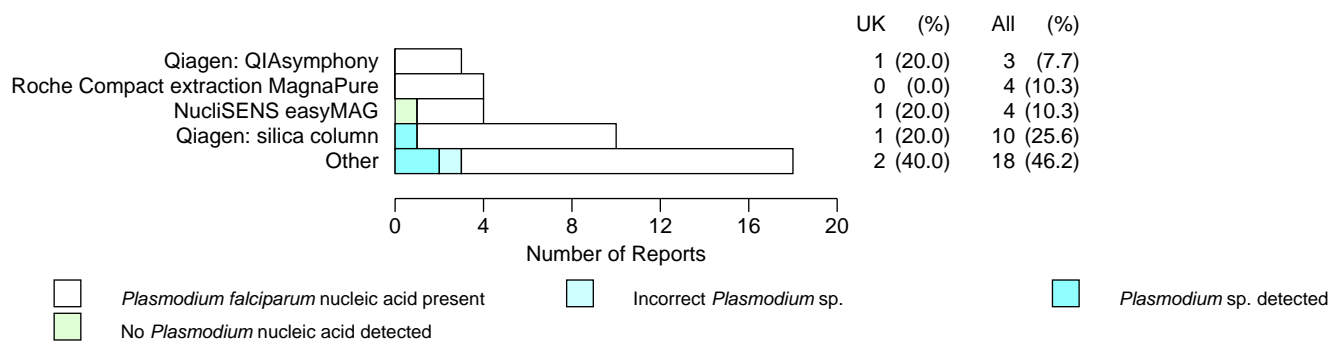
Specimen : 4463 *Plasmodium falciparum*: 200,000 parasites/mL **Detection Method 1**



Specimen : 4463 *Plasmodium falciparum*: 200,000 parasites/mL **Detection Method 2**



Specimen : 4463 *Plasmodium falciparum*: 200,000 parasites/mL **Extraction Methods**

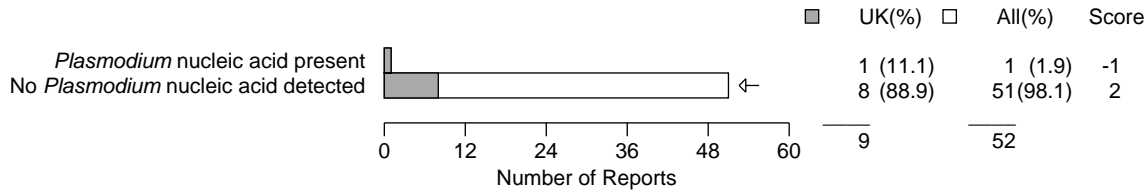


All specimens in this distribution were from a single patient sample and were diluted to the required parasite density using a donor blood which was negative for malaria DNA.

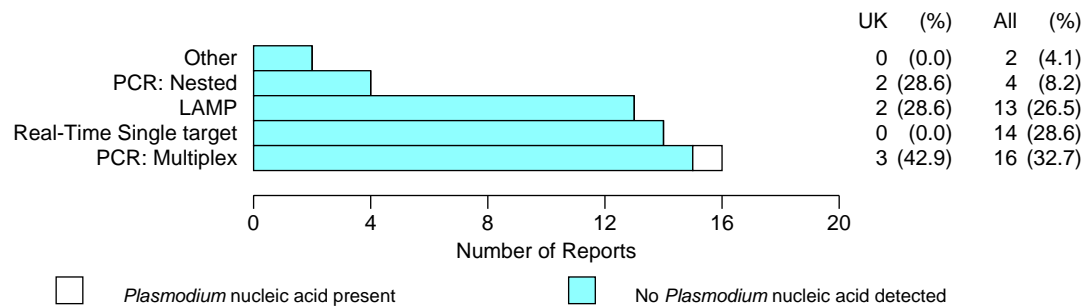
All samples were examined pre- and post- distribution by Nested PCR (modified Snounou protocol adapted from Snounou et al; 1993) and gave the intended results.

Please note that we report the parasitaemia in parasites/mL. Participants wanting to know the parasitaemia in parasites/uL are advised to divide the figure by 1000.

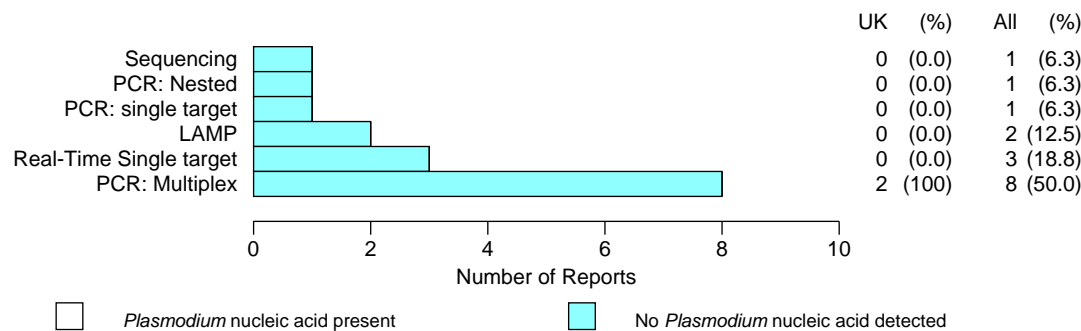
Specimen : 4464 *Plasmodium* species nucleic acid not present [Overall Results](#)



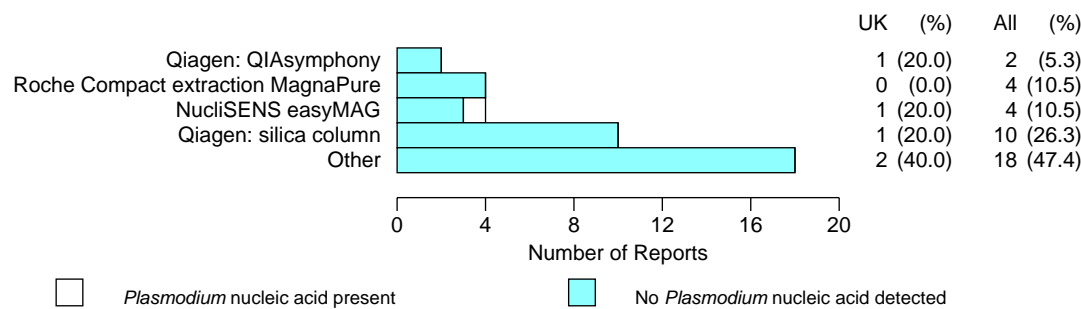
Specimen : 4464 *Plasmodium* species nucleic acid not present [Detection Method 1](#)



Specimen : 4464 *Plasmodium* species nucleic acid not present [Detection Method 2](#)



Specimen : 4464 *Plasmodium* species nucleic acid not present [Extraction Methods](#)

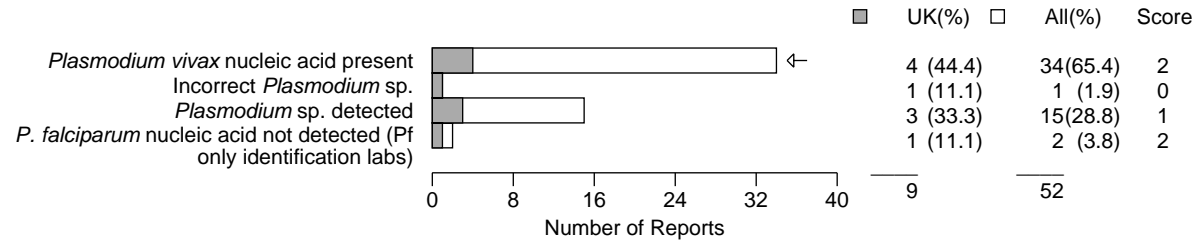


All specimens in this distribution were from a single patient sample and were diluted to the required parasite density using a donor blood which was negative for malaria DNA.

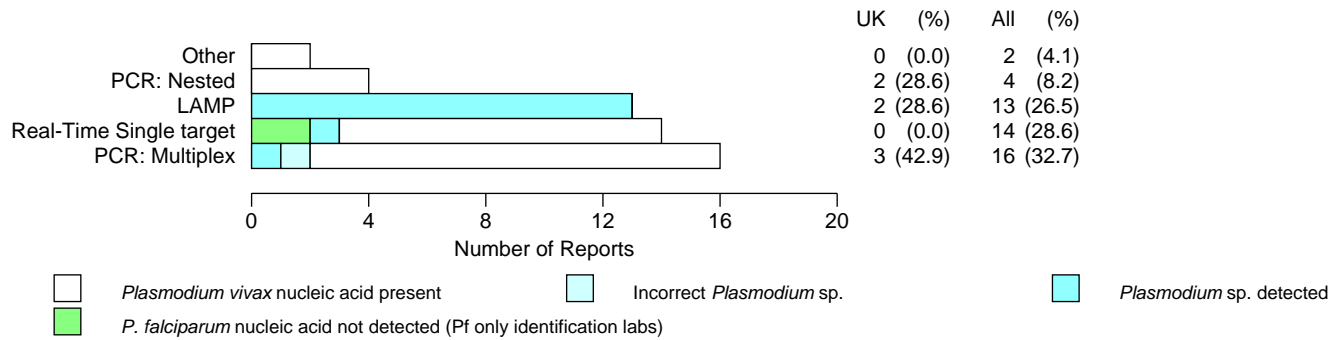
All samples were examined pre- and post- distribution by Nested PCR (modified Snounou protocol adapted from Snounou et al; 1993) and gave the intended results.

Please note that we report the parasitaemia in parasites/mL. Participants wanting to know the parasitaemia in parasites/uL are advised to divide the figure by 1000.

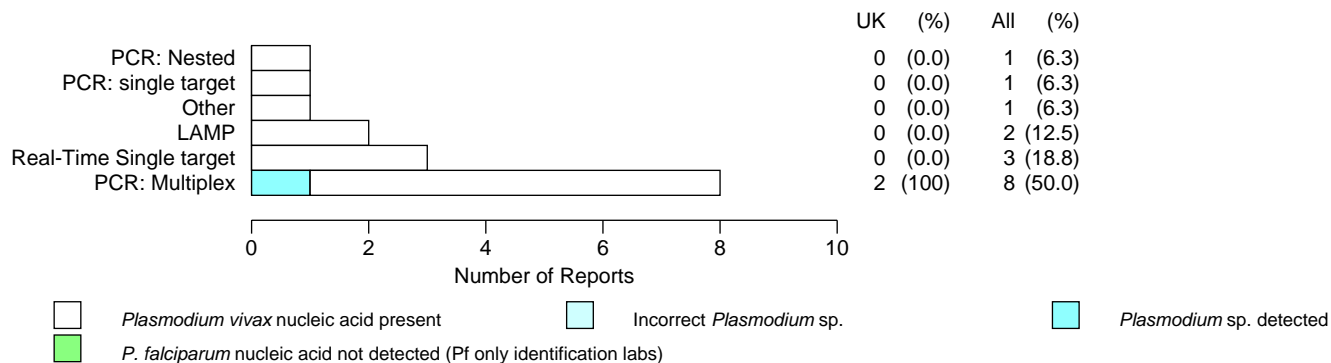
Specimen : 4465 *Plasmodium vivax*: 46,400 parasites/mL [Overall Results](#)



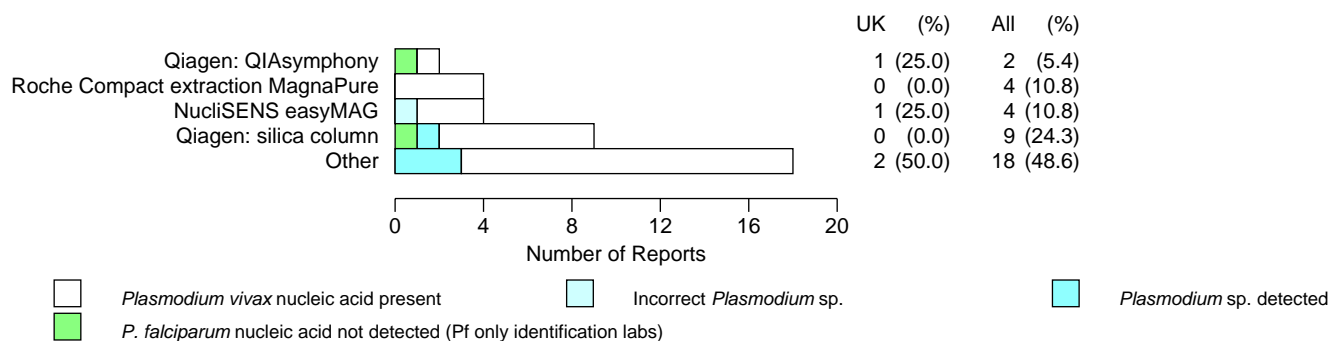
Specimen : 4465 *Plasmodium vivax*: 46,400 parasites/mL [Detection Method 1](#)



Specimen : 4465 *Plasmodium vivax*: 46,400 parasites/mL [Detection Method 2](#)



Specimen : 4465 *Plasmodium vivax*: 46,400 parasites/mL [Extraction Methods](#)

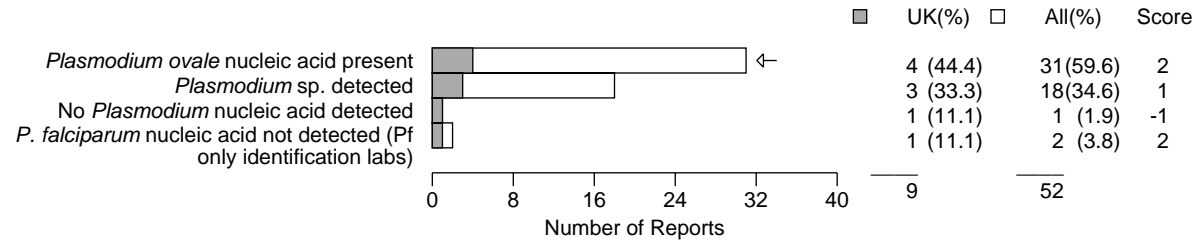


All specimens in this distribution were from a single patient sample and were diluted to the required parasite density using a donor blood which was negative for malaria DNA.

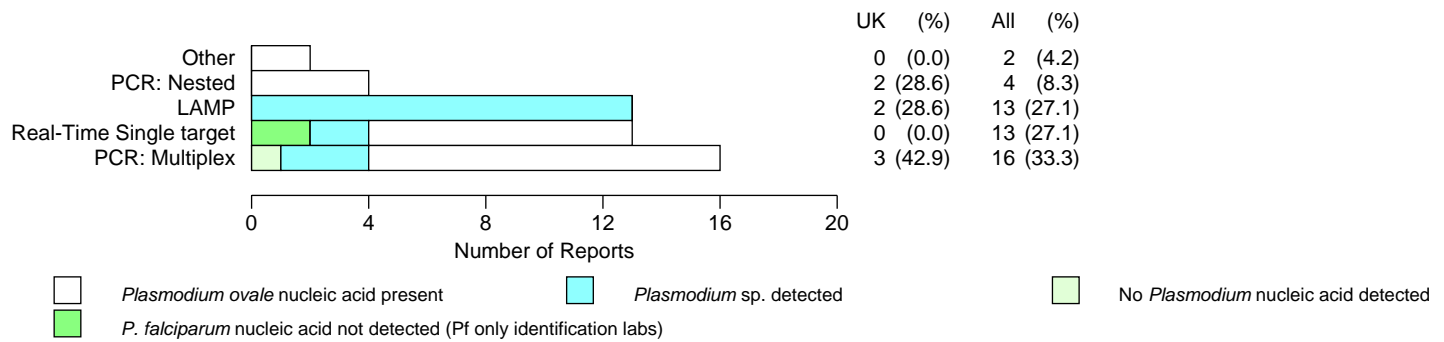
All samples were examined pre- and post- distribution by Nested PCR (modified Snounou protocol adapted from Snounou et al; 1993) and gave the intended results.

Please note that we report the parasitaemia in parasites/mL. Participants wanting to know the parasitaemia in parasites/uL are advised to divide the figure by 1000.

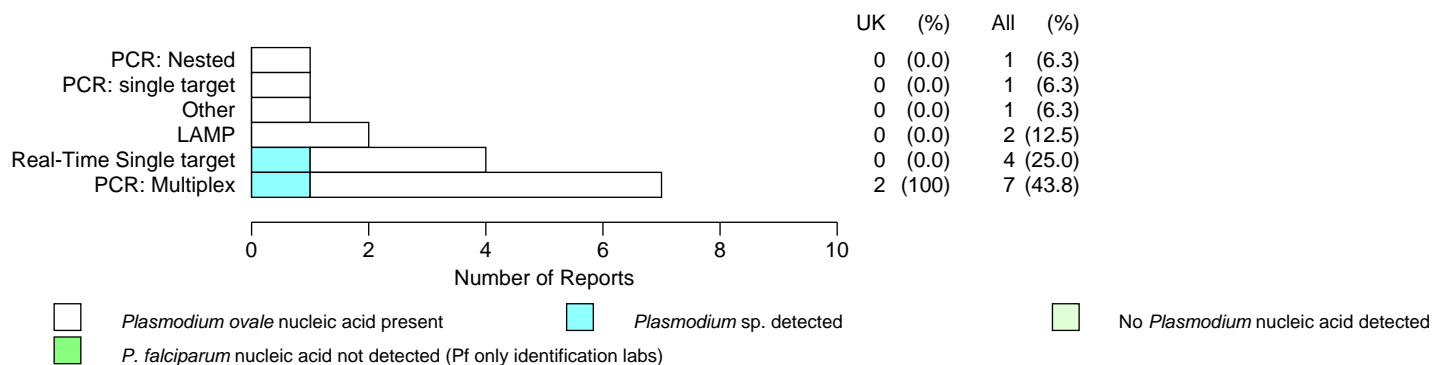
Specimen : 4466 *Plasmodium ovale*: 20,000 parasites/mL [Overall Results](#)



Specimen : 4466 *Plasmodium ovale*: 20,000 parasites/mL [Detection Method 1](#)



Specimen : 4466 *Plasmodium ovale*: 20,000 parasites/mL [Detection Method 2](#)



Specimen : 4466 *Plasmodium ovale*: 20,000 parasites/mL [Extraction Methods](#)

